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Response to Missing Parts/ Incomplete Application								
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SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT								
Firm BioMedPatent or John S. Sundsmo, Ph.D., 34,446								
Individual name Signature								
Date September 2, 2004								
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This collection of information is required by 37 CFR 1.5. The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to 2 hours to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants: Christian

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Agent Docket No. IMI-002

Serial No: 09/547,501

Group Art Unit: 1617

Filed:

April 12, 2000

Examiner: Shaojia A. Jiang

Title:

NOVEL PHARMACEUTICAL AGENTS CONTAINING CARBOHYDRATE MOIETIES AND METHODS OF THEIR

PREPARATION AND USE

SUPPLEMENTAL RESPONSE TO OFFICE ACTION

AND COURTESY COPY OF LIKHOSHERSTOV ET AL.

Vista, California 92085 September 2, 2004

TO THE COMMISSIONER OF PATENTS AND TRADEMARKS:

Office Action: Courtesy copy of Likhosherstov et al. and Supplemental response are now made in regard to the Office Action ("Action") carrying a mailing date of February 26, 2004 setting a 3 month period for response expiring May 26, 2004, i.e., extended with payment of fee under 37 C.F.R. § 1.136(a) to expire on August 26, 2004; and Request for Continued Examination filed with Response and Amendment on August 26, 2004.

<u>Documents Transmitted</u>: Copy of Likhosherstov et al. recently acquired from CNRS (Cedex, Paris, France); and, supplemental response.

Supplemental Response In Re. Likhosherstov et al.

With apologies to the Examiner, Applicant encountered some considerable difficulties in locating a copy of the Likhosherstov et al. article cited by the office as a Chem. Abstract. With the full text now received from France, some additional and clarifying response seems appropriate. A courtesy copy is transmitted herewith in the event that the Examiner may have encountered similar difficulties.

In regard to motivation in Likhosherstov et al., at page 1244 appears the following: namely,

"Scheme 1 Sug β 1-NHCOH₂Cl + HNR¹R² Sug β 1-NHCOCH₂NR¹R²

Sug = D-Gal(β 1-4)D-Gluc (a); D-GlcNAc (b); D-Gal (c); D-Man (d); D-Glc (e)" (page 1244, right column, lines 1-5; emphasis added);

"Alkylation of secondary amines was studied using morpholine and piperazine and its derivatives (N-methylpiperazine and 1,4-diazabicyclo[4.3.0]nonane). Piperazine is a well-known anthelminthic, and its structural fragment is a part of anesthetic, psychotropic, and antitumor drugs. Lectins specific to the residues of β -D-glucosamine are widespread on the surface of animal cells. Therefore, for the synthesis of glycoconjugates we used derivatives of these monosaccharides and the disaccharide lactose." (page 1244, right column, lines 8-17; emphasis added);

"The introduction of mono- and oligosacchearide residues into the molecules of various physiologically active compounds and drugs presents considerable interest due to the possibility of controlled change of their interactions with receptors and of target-directed transport to particular cells, which contain specific carbohydrate-binding proteins (lectins) on their surface." (page 1244, left column, lines 1-7; emphasis added).

Likhosherstov et al. teaches away from the claimed invention, i.e., the instant reaction product of claimed invention as compared with the disclosure above, is as follows: namely,

Sug-N-CR₁-Z-A (drug),

wherein R₁ and Z (when present) comprise lower alkyl or substituted lower alkyl.

Likhosherstov et al. motivates binding at \underline{N} -acetyl-glucos \underline{aminyl} -"lectins"/"receptors". Chemical properties required for binding at those receptors, i.e., the presence of \underline{N} -acetyl-glucosyl- \underline{N} H₂, are thus motivated by Likhosherstov et al. and not the requisite properties for blood brain barrier transport of glycosyl compounds. The difference makes a difference, in that (as related in prior responses and in the instnat Specification) sugar transporters are stereo- and anomer-specific and glucose, not glucosamine or NAc-glucosamine, is required for binding at glucose blood brain barrier transporters (GLUT). As related previously, even galactose (differing in anomeric position of

a hydroxyl) is not transported. Motivating different chemistry, thus, makes a considerable and significant difference in the expected outcome. Thus, different chemistry, different motivation, and expectation of different success are all features of Likhosherstov et al. Obviousness must be certain. Applicant believes Likhosherstov et al. teaches away.

Concluding Remarks

In light of the amendments to the claims and remarks transmitted August 26, 2004, as well as the supplemental remarks herein, removal of the rejections under 35 U.S.C. § 103 is respectfully requested. If any issues remain which can be expeditiously addressed in teleconference, the Examiner is urged to contact Applicant's agent at 760-806-3385 (office) or 615-423-3850 (mobile).

Respectfully submitted:

John S. Sundsmo, PhD Registration No.: 34,446

Glycoconjugates of amines: alkylation of primary and secondary amines with N-chloroacetyl- β -glycopyranosylamines

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Efficient monoalkylation of a series of primary and secondary amines was demonstrated with the use of N-chloroacetylglycosylamines derived from D-glucose, D-galactose, D-mannose, N-acetyl-D-glucosamine, and lactose. The reaction was shown to be useful for incorporation of carbohydrate residues into physiologically active compounds. Glycoconjugates of some derivatives of piperazine, 2-phenylethylamine, tryptamine, and important biogenic amines (norephedrine, octopamine, dopamine) were prepared.

Key words: glycoconjugates, N-chloroacetylglycosylamines, piperazine, 2-phenylethylamine, tryptamine, norephedrine, octopamine, dopamine.

The introduction of mono- and oligosaccharide residues into the molecules of various physiologically active compounds and drugs presents considerable interest due to the possibility of controlled change of their interaction with receptors and of target-directed transport to particular cells, which contain specific carbohydrate-binding proteins (lectins) on their surface.

Modification of the initial compounds with glycosylamine derivatives appears to be a promising approach to this type of construction of drug precursors ("prodrugs"). The glycosylamines can be easily prepared from either monosaccharides¹⁻⁴ or complex oligosaccharides including the products of cleavage of natural N-glycoproteins. 5-7 The use of glycosylamines for the synthesis of glycoconjugates generally involves their N-acylation and subsequent modification based on reactions of functional groups present in the acyl residue (see reviews^{8,9}).

A convenient variant of this approach is transformation of glycosylamines into N-haloacetylglycosylamines, 7,10,11 which are subsequently used to modify peptides and proteins at the SH groups. 10,11 The transformation of N-chloroacetylglycosylamines of oligosaccharides into N-glycylglycosylamines after the reaction with (NH₄)₂CO₃, and the use of the products for preparing various glycoconjugates containing fluorescent labels, biotin residues, palmitic acid, and also bovine serum albumin have been described. 7,12,13 However, the possibilities of using N-haloacetylglycosylamines for introduction of carbohydrate residues into other types of biologically active compounds still remain little studied.

In this work, we studied the reaction of some N-chloroacetyl-β-glycopyranosylamines 1a—e, described in our previous publication, with a number of primary and secondary amines (Scheme 1).

This reaction was used to alkylate a number of physiologically active compounds, which made it pos-

Scheme 1

Sugβ1-NHCOCH₂Cl + HNR¹R² → Sugβ1-NHCOCH₂NR¹R²
1a—e

Sug = p-Gal(β 1-4)p-Gic (a); p-GicNAc (b); p-Gal (c); p-Man (d); p-Gic (e)

sible to prepare previously unknown glycoconjugates 2-8 (Scheme 2).

Alkylation of secondary amines was studied using morpholine and piperazine and its derivatives (N-methylpiperazine and 1,4-diazabicyclo[4.3.0]nonane). Piperazine is a well-known anthelminthic, and its structural fragment is a part of anesthetic, psychotropic, and antitumor drugs. Lectins specific to the residues of β -D-galactose and N-acetyl-D-glucosamine are widespread on the surface of animal cells. ¹⁴ Therefore, for the synthesis of glycoconjugates we used derivatives of these monosaccharides and the disaccharide lactose.

We found that secondary amines are smoothly alky-lated upon treatment with chloroacetyl derivatives of glycosylamines (molar ratio 2:1) at 70 °C in MeOH or aqueous MeOH. The course of the reaction can be conveniently monitored by paper electrophoresis. After the reaction has been carried out for 3 h, the conversion of the alkylating reagent was 85—90%. Under the conditions chosen, we did not observe noticeable cleavage of the N-glycosylamide bond, whose lability in an alkaline medium has been noted previously for a glucose derivative. The reaction products were separated from the initial N-chloroacetylglycosylamines by the cation exchange chromatography and additionally purified by crystallization or chromatography on Al₂O₃; the yields of conjugates 2, 3c, and 4 were about 65%.

Translated from Izvestiya Akademii Nauk. Seriya Khimicheskaya, No. 6, pp. 1244-1247, June, 1998.

and dissolved in 2 mL of H₂O. The solution was applied to a column (1×13 cm) packed with the Amberlite 1RC-50 cation exchanger (H⁺). The column was washed with 100 mL of H₂O and 100 mL of 0.5 M aqueous pyridine. The fractions that contained product 2, according to the electrophoresis data, were combined and concentrated to dryness, and the residue was recrystallized from MeOH. Yield 0.29 g (63%), m.p. 214-216 °C, $[\alpha]_D^{20}$ +4.8° (c 1, H₂O). Found (%): C, 45.11; H, 7.11; N, 6.10; H_2O , 2.80. $C_{18}H_{32}N_2O_{12} \cdot 1/2$ H_2O . Calculated (%): C, 45.27; H, 6.96; N, 5.87; H₂O, 1.89. H NMR, δ: 2.62 (br.s, 4 H, CH₂NCH₂); 3.22 (br.s, 2 H, COCH₂); 3.42-3.97 (m, 16 H); 4.47 (d, 1 H, H(1) Gal, J=8 Hz); 5.03 (d, 1 H, H(1) Glc, J = 9 Hz). ¹³C NMR, 8: 54.5 (CH₂NCH₂); 61.7, 62.3, 62.9 (2 CH₂OH, COCH₂); 68.1 (CH₂OCH₂); 70.4, 72.8, 73.3, 74.4, 76.95, 77.2, 78.3, 79.6 (C(2)-C(5) Glc and Gal), 80.8 (C(1) Glc); 104.7 (C(1) Gal); 175.0 (CO).

N-Piperazinoacetyl-2-acetamido-2-deoxy-β-D-glucopyranosylamine (3a) and N-piperazinoacetyl-β-D-galactopyranosylamine (3b). Piperazine (0.87 g, 10 mmol) was added to a solution of N-chloroacetyl-2-acetamido-2-deoxy-β-D-glucopyranosylamine (1b) (0.3 g, 1 mmol) in 7 mL of MeOH or N-chloroacetyl-β-D-galactopyranosylamine (1c) (0.25 g, 1 mmol) in 12 mL of 70% aqueous MeOH. The mixture was heated for 3 h at 70 °C. The MeOH was evaporated, and the residue was dissolved in 20 mL of H_2O ; 35 mL of the Dowex 50Wx8 cation exchanger (H⁺) was added, and the mixture was stirred for 1 h. The resin was filtered off and washed with 400 mL of H_2O and 300 mL of 1.5 M NH₄OH. The alkaline fractions were concentrated to dryness, and the reaction products were isolated from the residue.

Compound 3a was isolated by chromatography on a column (2.8×11 cm) packed with Al_2O_3 (propan-2-ol \rightarrow MeOH) followed by crystallization (MeOH—propan-2-ol). Yield 0.2 g (58%), m.p. 218—219 °C, $[\alpha]_D^{20}$ +26.7° (c 1, H_2O). Found (%): C, 48.42; H, 7.55; N, 16.25. $C_{14}H_{26}N_4O_6$. Calculated (%): C, 48.54; H, 7.57; N, 16.17. H NMR, 8: 2.05 (s, 3 H, CH₃CO); 2.58 (br.s, 4 H, CH₂NCH₂); 2.92 (br.s, 4 H, CH₂NHCH₂); 3.20 (s, 2 H, COCH₂); 3.51—3.98 (m, 6 H); 5.15 (d, 1 H, H(1), J = 9 Hz).

Compound 3b was isolated by crystallization (MeOH—propan-2-ol). Yield 0.21 g (58%), m.p. 219—221 °C (decomp.), $[\alpha]_D^{20}$ +14.5° (c 1, H₂O). Found (%): C, 47.24; H, 7.63; N, 13.52. C₁₂H₂₃N₃O₆. Calculated (%): C, 47.20; H, 7.59; N, 13.76. ¹H NMR, 8: 2.60 (br.s, 4 H, CH₂NCH₂); 2.95 (br.s, 4 H, CH₂NHCH₂); 3.25 (br.s, 2 H, COCH₂); 3.66—3.87 (m, 5 H); 4.04 (m, 1 H, H(4)); 5.03 (d, 1 H, H(1), J = 9 Hz).

N-(4-Methylpiperazin-1-ylacetyl)-2-acetamido-2-deoxy-βp-glucopyranosylamine (3c). A mixture of N-chloroacetyl-2-acetamido-2-deoxy-β-D-glucopyranosylamine (1b) (0.45 g, 1.5 mmol) and N-methylpiperazine (0.3 mL, 3 mmol) in 5 mL of MeOH was heated for 3 h at 70 °C. The solvent was evaporated, and the residue was dissolved in 15 mL of H₂O; 12 mL of the Dowex 50Wx8 cation exchanger (H+) was added, and the mixture was stirred for 1.5 h. The resin was filtered off, washed with 200 mL of H₂O and then with 200 mL of 1.5 M NH₄OH. The alkaline fractions were evaporated to dryness. Recrystallization of the residue (MeOHacetone) gave compound 3c. Yield 0.4 g (74%), m.p. 205-207 °C, $[\alpha]_D^{20}$ +24.7° (c 1, H₂O). Found (%): C, 47.64; H, 8.08; N, 15.37; H₂O, 4.56. C₁₅H₂₈N₄O₆·H₂O. Calculated (%): C, 47.61; H, 7.99; N, 14.81; H₂O, 4.76. ¹H NMR, δ: 2.06 (s, 3 H, CH₃CO); 2.30 (s, 3 H, NCH₃); 2.57 (br.s. 8 H, 4 CH₂); 3.19 (br.s, 2 H, COCH₂); 3.50-3.96 (m, 6 H); 5.13 (d, 1 H, H(1), J = 9.5 Hz).

N-(2-Acetamido-2-deoxy-β-D-glucopyranosyl)-2-(1,4diazabicyclo[4.3.0]non-4-yl)acetamide (4). Compound 4 was prepared similarly to 3c from 1,4-diazabicyclo[4.3.0]nonane (0.375 g, 3 mmol)18 and purified by chromatography on a column (2.5×8 cm) packed with Al₂O₃ (propan-2-ol → MeOH) followed by crystallization (MeOH—ether). Yield 0.5 g (65%), m.p. 150-151 °C, $[\alpha]_D^{20}$ +21.7° (c 1, H₂O). Found (%): C, 50.40; H, 7.87; N, 14.52; H₂O, 4.56. C₁₇H₃₀N₄O₆·H₂O. Calculated (%): C, 50.48; H, 7.97; N, 13.85; H₂O, 4.45. ¹H NMR, δ: 1.38 (m, 1 H); 1.71-1.97 (m, 3 H); 2.02 (s, 3 H, CH₃CO); 2.05-2.47 (m, 5 H); 2.71-3.06 (m, 4 H); 3.20 (s, 2 H, COCH₂); 3.47-3.59 (m, 2 H, H(4,5) GlcN); 3.65 (t, 1 H, H(3) GlcN); 3.77 (dd, 1 H, H(6a) GlcN); 3.84-3.94 (m, 2 H, H(2b,6b) GlcN); 5.10 (d, 1 H, H(1) GlcN, J = 9.5Hz). ¹³C NMR, δ: 21.5 (CH₂); 23.1 (CH₃); 27.8 (CH₂); 51.2 (CH₂); 52.5 (CH₂); 53.3 (CH₂); 55.3 (C(2) GleN); 57.6 (CH₂); 61.3 (CH₂); 61.6 (CH₂); 63.1 (NCH); 70.7 (C(4) GlcN); 75.3 (C(3) GlcN); 78.8 (C(5) GlcN); 79.4 (C(1) GlcN); 174.6 (CO); 175.7 (CO).

N-(N-Phenethylglycyl)-2-acetamido-2-deoxy-β-D-glucopyranosylamine (5a) and N-(N-phenethylglycyl)-β-n-mannopyranosylamine (5b). 2-Phenylethylamine (0.4 mL, 3 mmol) and 10 mL MeOH were added to a solution of N-chloroacetyl-2-acetamido-2-deoxy-β-D-glucopyranosylamine (1b) (0.3 g, i mmol) or N-chloroacetyi-β-D-mannopyranosylamine (1d) (0.25 g, 1 mmol) in 2 mL of DMSO, and the mixture was heated for 10 h at 70 °C. The MeOH was evaporated, and the residue was diluted with 25 mL of toluene. The resulting oily product was washed 3 times with toluene and ether, and dissolved in 10 mL of H₂O. The solution was stirred for 1.5 h with 10 mL of the Dowex 50Wx8 cation exchanger (H⁺). The resin was filtered off, washed with 150 mL of H₂O, 150 mL of 1.5 M NH₄OH, and 150 mL of 1.5 M NH₄OH containing 6% Py. The alkaline fractions were concentrated to dryness, and the reaction products were isolated from the residue

Compound 5a was isolated by crystallization (MeOH—propan-2-ol). Yield 0.28 g (71%), m.p. 223—225 °C, $[\alpha]_D^{20}$ +21.4° (c 1, H₂O). Found (%): C, 55.41; H, 7.11; N, 11.02; H₂O, 2.12. C₁₈H₂₇N₃O₆·1/2 H₂O. Calculated (%): C, 55.37; H, 7.23; N, 10.76; H₂O, 2.31. ¹H NMR, δ: 1.95 (s, 3 H, CH₃CO); 2.84 (m, 4 H, CH₂CH₂); 3.34 (br.s, 2 H, COCH₂); 3.49—3.60 (m, 2 H, GlcN); 3.66 (t, 1 H, H(3) GlcN); 3.75—3.95 (m, 3 H, GlcN); 5.12 (d, 1 H, H(1) GlcN, J = 9.5 Hz); 7.32—7.50 (m, 5 H, Ar).

Compound 5b was isolated by chromatography on a column (1.4×12 cm) with Al₂O₃ (acetone \rightarrow propan-2-ol \rightarrow MeOH). The yield of the amorphous compound was 0.22 g (65%), $[\alpha]_D^{20}$ -24.8° (c 1, H₂O). Found (%): C, 56.71; H, 7.20; N, 8.23. C₁₆H₂₄N₂O₆. Calculated (%): C, 56.46; H, 7.11; N, 8.23. ¹H NMR, δ : 2.90 (m, 4 H, CH₂CH₂); 3.46 (br.s, 2 H, COCH₂); 3.48 (m, 1 H, H(5) Man); 3.62 (t, 1 H, H(4) Man); 3.68–3.78 (m, 2 H, Man); 3.88–3.98 (m, 2 H, Man); 5.24 (br.s, 1 H, H(1) Man); 7.30–7.48 (m, 5 H, Ar).

N-{N-[2-(Indol-3-yl)ethyl]glycyl}-β-D-galactopyranosylamine (6) was synthesized similarly to compound 5 from N-chloroacetyl-β-D-galactopyranosylamine (1c) (0.25 g, 1 mmol) and tryptamine (0.48 g, 3 mmol) in a mixture of 2 mL of DMSO and 18 mL of MeOH over a period of 30 h; prior to the treatment with the cation exchanger, the aqueous solution was decolorized by carbon. Product 6 was crystallized from H₂O. Yield 0.2 g (54%), m.p. 142–143 °C, $[\alpha]_D^{20}$ +15.4° (c 1, CH₃OH). Found (%): C, 54.90; H, 7.17; N, 10.58; H₂O, 3.55. C₁₈H₂₅N₃O₆·H₂O. Calculated (%): C, 54.40; H, 6.85; N, 10.57; H₂O, 4.53. ¹H NMR (CD₃OD), 8: 2.95 (br.s, 4 H, CH₂CH₂); 3.54–3.68 (m, 3 H, Gal);

3.71–3.78 (m, 2 H, Gal); 3.94 (m, 1 H, H(4) Gal); 4.92 (d, 1 H, H(1) Gal, J = 9 Hz); 6.99–7.15 (m, 2 H, Ar); 7.12 (s, 1 H, Ar); 7.37 (d, 1 H, Ar, J = 8 Hz); 7.58 (d, 1 H, Ar, J = 8 Hz). ¹³C NMR (CD₃OD), δ : 26.7 (CH₂Ar); 51.2 (NCH₂); 53.0 (NCH₂); 62.8 (C(6) Gal); 70.7 (C(4) Gal); 71.8 (C(2) Gal); 76.0 (C(3) Gal); 78.5 (C(5) Gal); 81.6 (C(1) Gal); 112.5, 113.8, 119.6, 119.8, 122.6, 123.8, 129.0, 138.5 (8 C, Ar); 175.4 (CO).

N-{N-[(1S,2R)-1-Hydroxy-1-phenylprop-2-yi]glycyl}-4-O- $(\beta-D-galactopyranosyl)-\beta-D-glucopyranosylamine (7a)$ and N-{N-[DL-2-hydroxy-2-(4-hydroxyphenyl)ethyl]glycyl}-4-O-(B-D-galactopyranosyl)-β-D-glucopyranosylamine (7b) was synthesized similarly to 5 from N-chloroacetyl-4-O-(β-D-galactopyranosyl)-β-D-glucopyranosylamine monohydrate (1a) (0.22 g, 0.5 mmol) and p-norephedrine hydrochloride (0.28 g, 1.5 mmol) or DL-octopamine hydrochloride (0.28 g, 1.5 mmol) in the presence of Et₃N (0.11 mL, 1.5 mmol) in a mixture of 1 mL of DMSO and 5 mL of MeOH. The reaction duration was 22 h. The residue obtained after concentration of the alkaline fractions resulting from ion exchange chromatography was treated with acetone (5×15 mL) in order to remove the initial amine (in the case of DL-octopamine, hot acetone was used) and dissolved in 5 mL of H2O. The solution was applied to a column (4×100 cm) with Sephadex G-25 (fine), and the column was washed with water (1 L) and then with 0.1 M AcOH. The fractions containing the products were combined, concentrated to dryness, dissolved in H2O, and lyophilized, and the amorphous residue was dried in vacuo over KOH.

Compound 7a was obtained in a yield of 0.16 g (61%), $[\alpha]_D^{20} +5.2^{\circ}$ (c 1, H_2O). Found (%): C, 51.42; H, 6.67; N, 5.23. $C_{23}H_{36}N_2O_{12}$. Calculated (%): C, 51.87; H, 6.81; N, 5.26. ¹H NMR, δ : 1.18 (d, 3 H, CH₃); 3.10 (m, 1 H, NCH); 3.48 (br.s, 2 H, COCH₂); 3.40—4.05 (m, 12 H, Glc, Gal); 4.55 (d, 1 H, H(1) Gal, J=8 Hz); 4.70 (d, 1 H, CHAr, J=6 Hz); 5.03 (d, 1 H, H(1) Glc, J=9 Hz); 7.46—7.58 (m, 5 H, Ar).

Compound 7b was obtained as the corresponding acetate by precipitation with ether from a solution in MeOH. Yield 0.21 g (79%) $[\alpha]_D^{20} + 4.5^\circ$ (c I, H₂O). Found (%): C, 48.10; H, 6.34; N, 4.51. C₂₁H₃₄N₂O₁₃·CH₃COOH. Calculated (%): C, 48.48; H, 6.44; N, 4.71. ¹H NMR, δ : 1.90 (s, 3 H, CH₃CO); 2.88 (m, 2 H, NCH₂); 3.38 (br.s, 2 H, COCH₂); 3.47—3.94 (m, 13 H); 4.44 (d, 1 H, H(1) Gal, J = 8 Hz); 4.98 (d, 1 H, H(1) Glc, J = 9 Hz); 6.89 (d, 2 H, Ar, J = 8 Hz); 7.32 (d, 2 H, Ar, J = 8 Hz).

 $N-\{N-[2-(3,4-\text{Dihydroxyphenyl})\text{ethyl}]\text{glyeyl}\}-\beta-D-gluco-pyranosylamine hydrochloride (8). Triethylamine (0.147 mL, 2 mmol) was added to a solution of <math>N$ -chloroacetyl- β -D-glucopyranosylamine (1e) (0.13 g, 0.5 mmol) and dopamine (0.38 g, 2 mmol) in a mixture of 1.3 mL of DMSO and 2.6 mL of MeOH. The mixture was heated for 2.5 h at 70 °C and poured in 45 mL of toluene, and the precipitate was washed with toluene (3×15 mL) and ether and dissolved in 3 mL of H_2O . To the solution, 4 mL of 0.5 H_2O (1.5 L) and the solution was applied to a column (5×90 cm) with Sephadex G-15. The column was washed with H_2O (1.5 L) and then with 0.1 H_2O (1.5 L) and then with 0.1 H_2O (1.5 L) and then products were combined, concentrated in vacuo, and lyophilized, and the amorphous residue was dried over H_2O . The yield of product 8 was 0.115 g (56%); H_2O 0 (c 1, H_2O 0).

Found (%): C, 46.60; H, 6.64; N, 6.23, Cl, 9.07. $C_{16}H_{24}N_2O_8 \cdot HCl$. Calculated (%): C, 47.00; H, 6.16; N, 6.85, Cl, 8.67. ¹H NMR, 8: 2.95 (t, 2 H, CH₂Ar, J = 7 Hz); 3.36 (t, 2 H, NCH₂, J = 7 Hz); 3.39—3.59 (m, 4 H, Glc); 3.73 (dd, 1 H, H(6a) Glc); 3.89 (dd, 1 H, H(6b) Glc); 3.98 (br.s, 2 H, COCH₂); 5.03 (d, 1 H, H(1), J = 9); 6.77 (d, 1 H, Ar, J = 8); 6.86 (s, 1 H, Ar); 6.42 (d, 1 H, Ar, J = 8).

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